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Mercury in Fish as a Potential Environmental Factor in the Development of Autoimmunity: A Mini-review with a Focus on Human Population Studies

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Abstract

Autoimmune diseases develop due to the interaction between genetic susceptibility and additional factors, such as environmental exposure to toxicants. Mercury (Hg), a well-established neurotoxin, has more recently been studied as an immunotoxin linked with biomarkers of autoimmunity, including the presence of antinuclear antibodies (ANA) and distinct cytokine profiles. Mercury (Hg) is virtually ubiquitous in the environment, and concerns about the potential health impacts of Hg exposure through fish consumption exist. A few studies have specifically examined the relationships among mercury, fish consumption, and autoimmune biomarkers in human populations. The findings of these studies are conflicting; this may be due to confounding exposures and opposing mechanisms of action. Additional studies are necessary to clarify the role of Hg through seafood consumption in autoimmunity.

Keywords: Autoimmune; Fish; Seafood; Mercury; Methyl mercury (MeHg); Antinuclear antibodies (ANA); Cytokines; N-3 PUFA; Selenium

Introduction

Autoimmune diseases develop due to interactions between genetic susceptibility and additional factors, including environmental exposure to toxicants [1]. Mercury (Hg) has been implicated as an environmental factor that contributes to the development and exacerbation of autoimmune disease [2]. Hg, a ubiquitous pollutant known to affect ecosystems and human health [3], exists in several chemical forms including inorganic mercury (iHg) and organic mercury (oHg). Microorganisms transform iHg present in sediment or water into oHg by methylation, yielding methyl mercury (MeHg). Plankton and algae absorb MeHg and are consumed by small fish, which are subsequently eaten by predators, ultimately resulting in biomagnification of MeHg up the food chain [4]. Humans are thus exposed to Hg through ingestion when they consume seafood into which Hg has bio accumulated, particularly because the form of MeHg in fish tissue is not removed through cooking or cleaning processes [4]. This mini-review discusses the potential autoimmune effects of MeHg with a focus on human MeHg exposure through fish consumption. This question of dietary MeHg exposure is significant because Hg is a global toxicant [3], and billions of people worldwide risk increased exposure to MeHg through reliance on fish as a major source of dietary protein and nutrition [5,6].

MeHg Immune Effects in Animal and *in vitro* Studies

Animal models provide evidence for Hg's role in inducing autoimmune effects. Exposing genetically-susceptible mouse strains to Hg leads to the development and/or exacerbation of lupus-like symptoms [7-12], including increased antinuclear autoantibodies (ANA) [7,8]. MeHg exposure in mice led to an initial immunosuppression via reduction in T- and B-cell populations [13], followed by an increase in ANA and IL-4 mRNA expression [13-15]. *In vitro* studies in human peripheral blood mononuclear cells (PBMCs) treated with sub-toxic MeHg resulted in increased concentrations of cytokine IL-1 β [16] and suppression of cytokines IL-2 and TGF- β [17]. These results support that MeHg, the form of Hg in human dietary sources, leads to immune dysregulation and autoimmunity. The preservation of food has several objectives [1].

MeHg, Fish Consumption, and Immune System Effects in Human Population Studies

Few human studies explicitly examine the role of Hg exposure through fish consumption with biomarkers of autoimmunity. Silva et al. [18] reports increased prevalence of ANA (10.7%) and antinucleolar antibodies (ANoA) (18%) in a population exposed to MeHg through fish consumption versus the reference site (ANA 7.1%, ANoA 2.0%), though the prevalence was not as elevated as those measured in occupationally exposed miners

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(54.1% ANA, 40.8% ANoA). Another study of Amazonian communities [19] observed positive serum ANA more frequently in riverine who consumed fish daily (including species with confirmed high MeHg) than in controls (12.4% vs. 2.9%), and mean hair Hg of riverine (34.5 ppm) was significantly higher than controls (1.0 ppm). Despite the significant differences in both ANA and mean hair Hg in riverine versus control communities, there was no significant association between hair Hg and ANAs [19]. In a similar trend, our study [20-23] of participants residing on Cheyenne River Sioux Tribe (CRST) Lands, a known area of MeHg contamination, found a relationship between fish consumption and elevated levels of ANA and specific autoantibodies, yet blood Hg itself was not associated with autoantibodies. In both Amazonian Brazil and CRST studies, fish consumption, but not Hg measure in biological matrices, is associated with increased autoantibodies.

In seeming contrast, more recent studies published on Hg and autoimmune biomarkers in Hg-exposed populations in Columbia [24] and the Middle Atlantic Coast of the United States (Long Island) [25] show that fish consumption is significantly associated with increased levels of Hg in biological matrices, yet these Hg measures are not associated with altered levels of ANAs [24,25], rheumatoid factors (RF) [24], or cytokines [25]. It is difficult to isolate the effects of Hg exposure from eating fish with the effects of Hg exposure from other sources because Hg is an environmentally pervasive contaminant, and people who regularly consume Hg-contaminated local fish likely also encounter Hg through occupational or other environmental exposures. In the case of the studies in Amazonian Brazil [18,19] and Columbia [24], additional iHg exposures result from gold mining, while the studies of CRST [23] and Long Island [25] cite iHg exposures from emissions and industry. Although all studies discussed in this mini-review [18-25] show an increase in total Hg in biological blood and/or hair correlated specifically to reported fish consumption, only the Silva et al. Amazonian Brazil [18] implies a full linkage from fish consumption to increased concentration of bodily Hg, and Hg body burden with increased autoimmune markers.

Discussion

The fact that fish consumption is associated with autoimmune markers in Amazonian Brazil [18,19] and CRST [23] studies may be due to additional exposures to contaminants implicated in immune dysregulation. Fish consumption likely serves as an exposure surrogate or composite exposure predictor. Participants in these studies reside in environments impacted by mine wastes that include other metals (gold, cadmium, arsenic) known to play a role in autoimmunity [26-28]. Additionally, pesticide exposure was not adjusted for in the Amazonian Brazil [18,19] and CRST [23] studies. Like Hg, pesticides are persistent environmental contaminants capable of bio accumulating in fish and have been implicated in immune alterations [29]. The adjustment for pesticide exposure may explain the lack of autoantibody induction observed in the Columbia study [24] in spite of the fact that this population also resides in a gold mining setting.

Differences in genetic, metabolic, lifestyle and total environmental exposure across populations are also likely contributors to the discrepancies in findings. A notable difference among these human studies is the total body burden of Hg in the study populations. The CRST, Columbian, and Long Island population studies all measured low levels of total blood Hg in comparison to the Amazonian Brazil studies, and no significant associations between total Hg and autoimmune markers were observed. This suggests that chronically high total body burden of Hg, rather than MeHg from fish, is associated with increased autoimmune markers. This idea is supported by additional studies published on Amazonian Brazil mining communities without reported fish consumption that showed positive associations between high total hair Hg and ANA, ANoA, and cytokines (IL-1 β , TNF- α , IFN- γ) [16-22]. An alternative possibility to a minimum total Hg exposure, or an additional requirement, may be that effective induction of autoimmune markers requires the presence of both iHg and MeHg. iHg and MeHg have been shown in mice [26] and human PBMCs [16] to elicit differential immune responses with iHg favouring a Th2 response whereas MeHg favours a Th1 response. Furthermore, studies of Amazonian Brazil populations reported a high prevalence of malaria [16-22], which has been shown in mouse models to lead to the generation of antibodies that react with nuclear antigens [27]. This suggests that a convergence of factors; iHg, MeHg, and specific immune challenge, such as malaria infection increases the probability of autoimmunity.

Other than exposures to additional environmental contaminants, selenium (Se), and fatty acids consumed alongside MeHg in fish may account for some of the uncertainty in the associations between Hg-contaminated fish consumption and autoimmunity. A study of Hg miners in China who had correlated elevated Hg and Se found increased selenoproteins and glutathione peroxidase (GSH-Px) [30], which may mitigate the adverse effects of Hg exposure contributing to the development of autoimmunity. The principle source of Se is through dietary animal protein [31], and some authors' state that Se, like MeHg, biomagnifies within predatory fish [32]. Others suggest that Se accumulates at the base of the food chain and that significant concentrations of Se may be ingested through plants grown in a Se-enriched environment [33]. A follow-up study in Amazonian Brazil found an inverse relationship between blood Hg and blood Se but no overall relationship between fish consumption and Se even though fish consumption was high [34]. Although the primary source of Se intake is unclear, Se and Hg are correlated in both the environment and the human body, and there is evidence that they have opposing mechanisms of action.

n-3 Poly-unsaturated fatty acids (n-3 PUFA) present in fish may also counteract the negative effects of Hg on the immune system. n-3 PUFAs are known to have the ability to regulate transcription factor activation and pro-inflammatory signalling pathways and may modulate pathways involved in autoimmune disorders [34-36]. This potentially explains why the Long Island study [25], the only one to measure n-3 PUFAs in participants, found correlations between n-3 PUFAs and detection of ANA only at lower titre concentrations.

Finally, it is worth mentioning that common markers of autoimmunity such as ANA and ANoA are generally observed at low frequency at the conservative titres (1:80 or more dilute) used in the human studies cited here, and many cytokine measurements lie below the limit of detection. This, and limited population sample size, pose additional obstacles to reaching a firm conclusion about the role of MeHg fish in the development of autoimmune biomarkers.

Conclusions and Future Avenues of Study

The findings of the few human studies incorporating MeHg exposures through fish consumption do not provide a conclusive answer as to whether or not these exposures significantly contribute to autoimmune development. In our study with the CRST [23], which exhibits elevated levels of certain autoimmune diseases, the main question from community members was, "Is it safe to eat local fish?" Reframed, the question is, "Does MeHg fish consumption exacerbates the development from autoimmunity?" Current studies do not provide a clear consensus. It appears that high total Hg body burden is necessary in order to observe significant changes in autoimmune biomarkers. A combination of both iHg and MeHg exposures may be required to exacerbate autoimmune development, since the various forms of Hg affect the immune system differently. Because Amazonian Brazil populations evidenced both relatively high total Hg and increased likelihood of exposure to malaria, it is possible that development of Hgdriven autoimmunity in humans depends upon a convergence of factors: iHg, MeHg, and specific immune challenge, such as malaria infection. It is likely that nutritional elements in fish, including Se and n-3 PUFAs, attenuate the immune effects of Hg exposures. The limited evidence in human populations about the role of fish MeHg in autoimmunity concurs with the current public health consensus to retain or increase fish consumption, especially of species with lower MeHg, for nutritional benefits while decreasing other exposures to Hg.

To elucidate the question of whether or not MeHg through fish consumption contributes significantly to alterations in autoimmune markers in humans, a larger, more robust set of human studies is needed. Autoimmune biomarkers could be measured in populations exposed to MeHg through fish consumption, beginning with the many communities world-wide in which Hg bio monitoring in fish tissue and/or human biological samples has already been done [37-48]. Estimated MeHg exposure, calculated from accurate species-specific tissue MeHg concentrations, should be modelled as a predictor alongside measures of iHg exposure with autoimmune biomarkers as the outcomes. This would help disentangle fish consumption's role in autoimmunity from that of other Hg exposures in order to inform public health recommendations.

References

- 1. Heward J, Gough SC (1997) Genetic susceptibility to the development of autoimmune disease. Clin Sci 93: 479-491.
- Bagenstose LM, Salgame P (1999) Monestier M. Murine mercuryinduced autoimmunity. Immunol Res 20: 67-78.

- Driscoll CT, Mason RP, Chan HM, Jacob DJ, Pirrone N (2013) Mercury as a global pollutant: sources, pathways, and effects. Environ Sci Technol 47: 4967-4983.
- 4. Li P, Feng X, Qiu G (2010) Methylmercury exposure and health effects from rice and fish consumption: A review. Int J Environ Res Public Health 7: 2666-2691.
- 5. Ha E, Basu N, Bose-O'Reilly S, Dórea JG, McSorley E, et al. (2017) Current progress on understanding the impact of mercury on human health. Environ Res 152: 419-433.
- Mergler D, Anderson HA, Chan LHM, Mahaffey KR, Murray M, et al. (2007) Methylmercury exposure and health effects in humans: a worldwide concern. Ambio 36: 3-11.
- 7. Pollard KM, Hultman P, Kono DH (2005) Immunology and genetics of induced systemic autoimmunity. Autoimmun Rev 4: 282-288.
- Germolec D, Kono DH, Pfau JC, Pollard KM (2012) Animal models used to examine the role of the environment in the development of autoimmune disease: findings from an NIEHS Expert Panel Workshop. J Autoimmun 39: 285-293.
- Havarinasab S, Hultman P (2006) Alteration of the spontaneous systemic autoimmune disease in (NZB x NZW) F1 mice by treatment with thimerosal (ethyl mercury). Toxicol Appl Pharmacol 214: 43-54.
- Hultman P, Taylor A, Yang JM, Pollard KM (2006) The effect of xenobiotic exposure on spontaneous autoimmunity in (SWR x SJL)F1 hybrid mice. J Toxicol Environ Health A 69: 505-523.
- 11. Pollard KM, Pearson DL, Hultman P, Deane TN, Lindh U, et al. (2001) Xenobiotic acceleration of idiopathic systemic autoimmunity in lupus-prone bxsb mice. Environ Health Perspect 109: 27-33.
- 12. Pollard KM, Pearson DL, Hultman P, Hildebrandt B, Kono DH (1999) Lupus-prone mice as models to study xenobiotic-induced acceleration of systemic autoimmunity. Environ Health Perspect 107: 729-735.
- Häggqvist B, Havarinasab S, Björn E, Hultman P (2005) The immunosuppressive effect of methylmercury does not preclude development of autoimmunity in genetically susceptible mice. Toxicology 208: 149-164.
- 14. Havarinasab S, Björn E, Nielsen JB, Hultman P (2007) Mercury species in lymphoid and non-lymphoid tissues after exposure to methyl mercury: Correlation with autoimmune parameters during and after treatment in susceptible mice. Toxicol Appl Pharmacol 221: 21-8.
- 15. Hultman P, Hansson-Georgiadis H (1993) Methyl mercury-induced autoimmunity in mice. Toxicol Appl Pharmacol 154: 203-211.
- Gardner RM, Nyland JF, Silbergeld EK (2010) Differential immunotoxic effects of inorganic and organic mercury species in vitro. Toxicol Lett 198: 182-190.
- 17. Das K, Siebert U, Gillet A, Dupont A, Di-Poï C, et al. (2006) Mercury immune toxicity in harbour seals: links to in vitro toxicity. Environ Health 7: 52.
- Silva IA, Nyland JF, Gorman A, Perisse A, Ventura AM, et al. (2004) Mercury exposure, malaria, and serum antinuclear/antinucleolar antibodies in Amazon populations in Brazil: a cross-sectional study. Environ Health 3: 11.
- Alves MFA, Fraiji NA, Barbosa AC, De Lima DSN, Souza JR, et al. (2006) Fish consumption, mercury exposure and serum antinuclear antibody in Amazonians. Int J Environ Health Res 16: 255-262.

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- 20. Gardner RM, Nyland JF, Silva IA, Ventura AM, De JM, et al. (2011) 35 Mercury exposure, serum antinuclear/antinucleolar antibodies and serum cytokine levels in mining populations in Amazonian
- 21. Nyland JF, Fillion M, Barbosa F, Shirley DL, Chine C, et al. (2011) Biomarkers of methylmercury exposure immunotoxicity among fish consumers in Amazonian Brazil. Environ Health Perspect 119: 1733-1738.

Brazil: A cross-sectional study 110: 345-354.

- 22. Motts JA, Shirley DL, Silbergeld EK, Nyland JF (2014) Novel biomarkers of mercury-induced autoimmune dysfunction: A cross-sectional study in Amazonian Brazil. Environ Res 132: 12-18.
- 23. Ong J, Erdei E, Rubin RL, Miller C, Ducheneaux C, et al. (2014) Mercury, autoimmunity, and environmental factors on Cheyenne River Sioux Tribal lands. Autoimmune Dis 2014: 325461.
- 24. Sánchez Rodríguez LH, Flórez-Vargas O, Rodríguez-Villamizar LA, Vargas Fiallo Y, Stashenko, et al. (2015) Lack of autoantibody induction by mercury exposure in artisanal gold mining settings in Colombia: Findings and a review of the epidemiology literature. J Immunotoxicol 12: 368-375.
- 25. Monastero RN, Karimi R, Nyland JF, Harrington J, Levine K, et al. (2017) Mercury exposure, serum antinuclear antibodies, and serum cytokine levels in the Long Island Study of Seafood Consumption: A cross-sectional study in NY, USA. Environ Res 156: 334-340.
- 26. Hultman P, Hansson-Georgiadis H (1999) Methyl mercury-induced autoimmunity in mice. Toxicol Appl Pharmacol 154: 203-211.
- Mannoor K, Li C, Inafuku M, Taniguchi T, Abo T, et al. (2013) Induction of ssDNA-binding autoantibody secreting B cell immunity during murine malaria infection is a critical part of the protective immune responses. Immunobiology 218: 10-20.
- Rowley B, Monestier M (2005) Mechanisms of heavy metalinduced autoimmunity. Mol Immunol 42: 833-838.
- 29. Mokarizadeh A, Faryabi MR, Rezvanfar MA, Abdollahi M (2015) A comprehensive review of pesticides and the immune dysregulation: mechanisms, evidence and consequences. Toxicol Mech Methods 25: 258-278.
- Chen C, Yu H, Zhao J, Li B, Qu L, et al. (2006) The roles of serum selenium and selenoproteins on mercury toxicity in environmental and occupational exposure. Environ Health Perspect 114: 297-301.
- 31. Levander OA (1987) A Global View of Human Selenium Nutrition 7: 227-2250.
- 32. Barwick M, Maher W (2003) Biotransference and biomagnification of selenium copper, cadmium, zinc, arsenic and lead in a temperate seagrass ecosystem from Lake Macquarie Estuary, NSW, Australia. Mar Environ Res 56: 471-502.
- Chen Y-W, Belzile N, Gunn JM (2001) Antagonistic effect of selenium on mercury assimilation by fish populations near Sudbury metal smelters? Limnol Oceanogr 46: 1814-1818.
- Lemire M, Mergler D, Fillion M, Passos CJS, Guimarães JRD, et al. (2006) Elevated blood selenium levels in the Brazilian Amazon. Sci Total Environ 366: 101-111.

- 35. Simopoulos AP (2002) Omega-3 Fatty Acids in Inflammation and Autoimmune Diseases. J Am Coll Nutr 21: 495-505.
- 36. Calder PC (2010) Omega-3 fatty acids and inflammatory processes. Nutrients 2: 355-374.
- Gill R, Lanni L, Jen KLC, McCabe MJ, Rosenspire A (2015) Docosahexaenoic acid counteracts attenuation of CD95-induced cell death by inorganic mercury. Toxicol Appl Pharmacol 282: 61-67.
- Gill R, Jen KL, McCabe MJJ, Rosenspire A (2016) Dietary n-3 PUFAs augment caspase 8 activation in Staphylococcal aureus enterotoxin B stimulated T-cells. Toxicol Appl Pharmacol 309: 141-148.
- 39. Langeland AL, Hardin RD, Neitzel RL (2017) Mercury levels in human hair and farmed fish near artisanal and small-scale gold mining communities in the madre de dios River Basin, Peru. Int J Environ Res Public Health 14: E302.
- Miklavčič A, Casetta A, Snoj Tratnik J, Mazej D, Krsnik M, et al. (2013) Mercury, arsenic and selenium exposure levels in relation to fish consumption in the Mediterranean area. Environ Res 120: 7-17.
- 41. Dellinger JA (2004) Exposure assessment and initial intervention regarding fish consumption of tribal members of the Upper Great Lakes Region in the United States. Environ Res 95: 325-340.
- Rothschild RFN, Duffy LK (2002) Preliminary study on total mercury in the common prepared subsistence foods of a rural Alaskan village. Alaska Med 44: 89-93.
- 43. Shao D, Kang Y, Cheng Z, Wang H, Huang M, et al. (2013) Hair mercury levels and food consumption in residents from the Pearl River Delta: South China. Food Chem 136: 682-688.
- 44. Morrissette J, Takser L, St-Amour G, Smargiassi A, Lafond J, et al. (2004) Temporal variation of blood and hair mercury levels in pregnancy in relation to fish consumption history in a population living along the St. Lawrence River. Environ Res 95: 363-374.
- 45. Hsiao H-W, Ullrich SM, Tanton TW (2011) Burdens of mercury in residents of Temirtau, Kazakhstan I: hair mercury concentrations and factors of elevated hair mercury levels. Sci Total Environ 409: 2272-2280.
- 46. Guentzel JL, Portilla E, Keith KM, Keith EO (2007) Mercury transport and bioaccumulation in riverbank communities of the Alvarado Lagoon System, Veracruz State, Mexico. Sci Total Environ 388: 316-324.
- 47. Pataranawat P, Parkpian P, Polprasert C, Delaune RD, Jugsujinda A (2007) Mercury emission and distribution: Potential environmental risks at a small-scale gold mining operation, Phichit Province, Thailand. J Environ Sci Health A Tox Hazard Subst Environ Eng 42: 1081-1093.
- 48. You CH, Kim BG, Jo EM, Kim GY, Yu BC, et al. (2012) The relationship between the fish consumption and blood total/ methyl-mercury concentration of costal area in Korea. Neurotoxicology 33: 676-682.